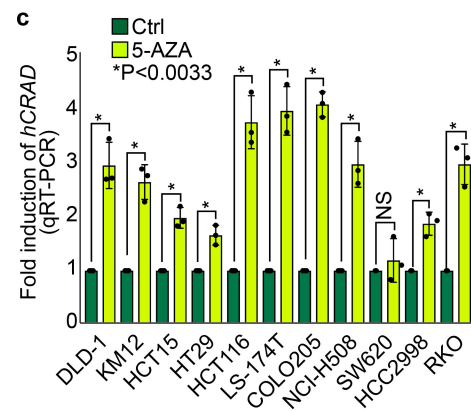
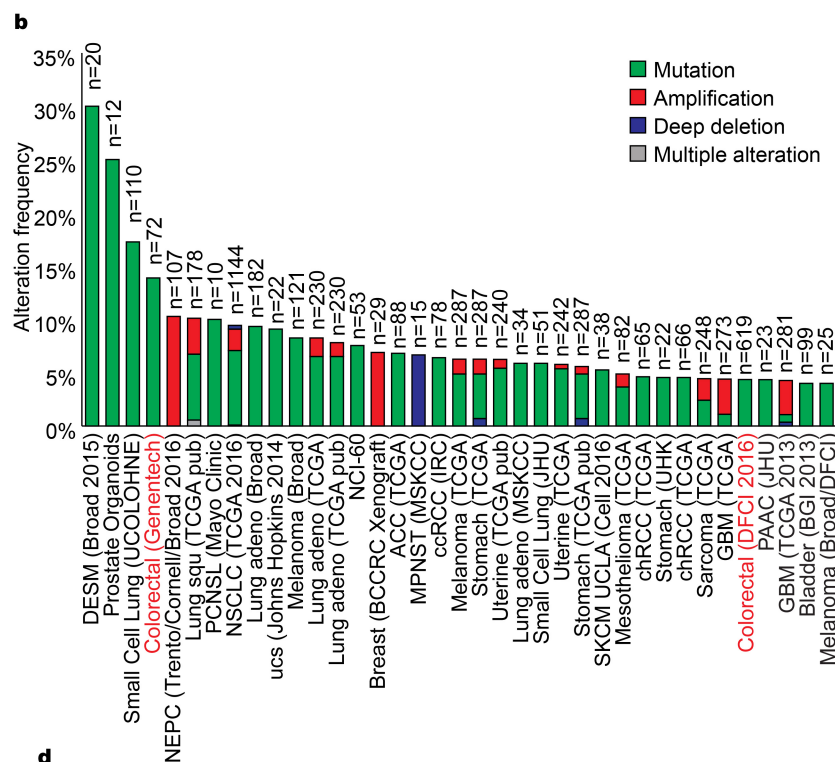


a

	<i>CRAD</i>		<i>APC</i>		<i>DCC</i>		<i>CTNNB1</i>		<i>AXIN2</i>		<i>HPRT1</i>	
Mutation	N	%	N	%	N	%	N	%	N	%	N	%
Substitution nonsense	5	3.18	1177	35.90	33	11.50	17	0.37	3	3.61	0	0
Substitution missense	97	61.78	494	15.07	207	72.13	3970	86.51	53	63.86	11	84.62
Substitution synonymous	38	24.20	171	5.22	60	20.91	90	1.96	13	15.66	3	23.08
Insertion inframe	11	7.01	3	0.09	0	0	6	0.13	0	0	0	0
Insertion frameshift	5	3.18	583	17.78	0	0	1	0.02	8	9.64	0	0
Deletion inframe	1	0.64	0	0	0	0	353	7.69	0	0	0	0
Deletion frameshift	6	3.82	1186	36.17	4	1.39	16	0.35	9	10.84	0	0
Nonsense + Frameshift (%)	10.18		89.85		12.09		0.74		24.09		0	



d

Cell line	Mutation	Cell line	Mutation	Cell line	Mutation
LIM2551	p.E309fs*115 (D)	LIM2405	p.Q324R (M)	RKO	p.A783T (M)
LIM2551	p.Q760fs*69 (D)	SW620	p.A501G (M)	HDC101	p.D15Y (M)
CCK81	p.K167fs*82 (D)	SW948	p.S975P (M)	SW48	p.A286V (M)
LIM1899	p.E309fs*115 (D)	HCT116	p.A511V (M)		

10 of 130 cell lines; M: Substitution-Missense; D: Deletion-Frame-shift

Supplementary Figure 1

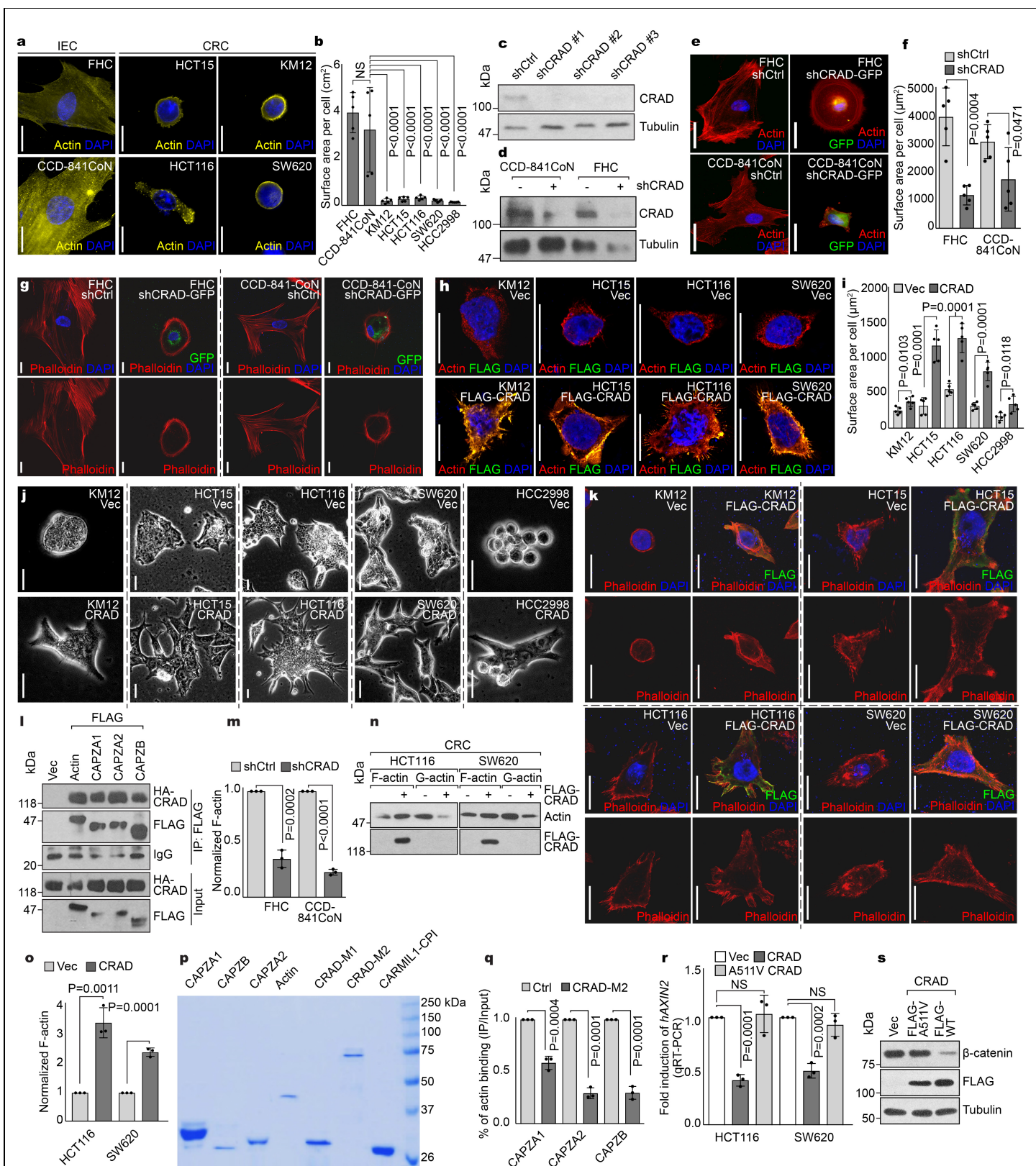
Genetic inactivation of *CRAD* in cancer

a, Genetic inactivation of *CRAD* in CRC. cBioportal database analysis of *CRAD* in CRC. *CRAD* shows 10.18 % incidence rate of truncated mutants (Nonsense + Frameshift) in CRC. *APC*, *DCC*, *CTNNB1*, and *AXIN2* were analyzed as the positive control. *HPRP1* served as negative control. Total 502 CRC cases were analyzed.

b, Genetic alteration of *CRAD* in human cancer. cBioportal analysis.

c, Potent epigenetic suppression of *CRAD* promoter. Upregulated *CRAD* expression by the inhibition of methylation. IECs and CRC cells were treated with 5-Azacytidine (5-AZA; 20μM) for 24hr. n= 3 independent experiments; Error bars: average ± S.D.; NS: not significant (P>0.05); Two-sided unpaired *t*-test.

d, COSMIC analysis of *CRAD* mutations in CRC cell lines.



Supplementary Figure 2

Positive regulation of the actin polymerization by CRAD-induced capping protein inhibition

a and b, Morphological comparison of IECs and CRC cells. IF staining (**a**). Quantification of the cell area of IECs and CRC cells by AxioVision software (**b**; n= 5 independent experiments)

c and d, Depletion of endogenous CRAD by shRNAs. FHC cells were transfected with shRNAs (**c**). shCRAD#1 stably expressing IECs were analyzed for IB (**d**). IB was performed once.

e-g, Cell shrinkage by CRAD depletion. 48hr after transfection, cells were analyzed for IF staining (**e**) and cell area quantification (**f**; n=5 independent experiments). Cells were infected with lenti-shCRAD for 48hr. After infection, cells were selected by puromycin (2µg/ml) treatment for 72hr. The cell morphology was analyzed by IF staining for Phalloidin (**g**).

h-k, Extended cell morphology by CRAD in CRC cells. 48hr after transfection (Vec [empty vector] or FLAG-CRAD), cells were analyzed for IF staining (actin, **h**; Phalloidin, **k**) and cell area quantification (**i**; n=5 independent experiments). The cell morphology was monitored using bright field microscope (**j**).

l, Physical interaction of CRAD with CPs. Purified FLAG-Actin, FLAG-CPs, and HA-CRAD proteins were used for co-IP. These results recapitulate *in vitro* interaction between CRAD and CPs performed using GST recombinant proteins. Experiment was performed three times with similar results.

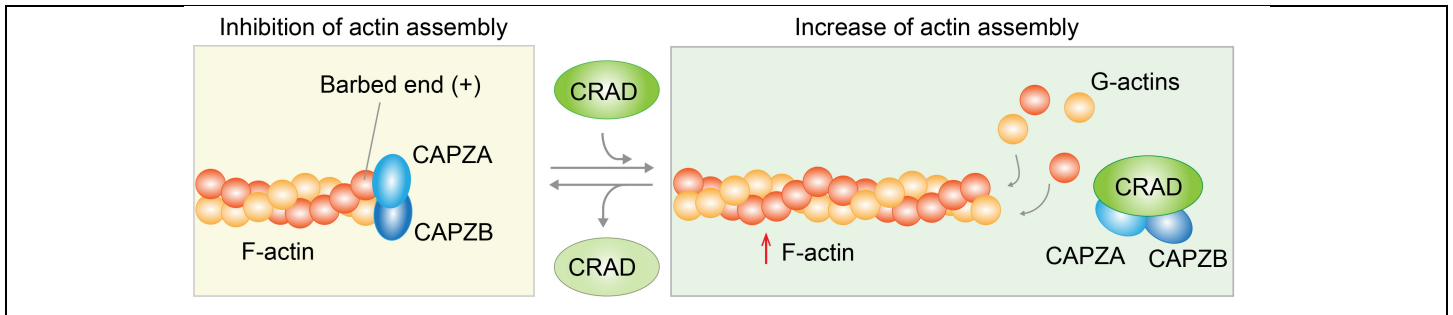
m-o, Decreased F-actin formation by CRAD depletion. Quantification of the F-actin in the CRAD depleted (**m**), or the CRAD ectopic expressed condition (**n and o**). The level of F-actin and G-actin was examined by immunoblots (**n**) and quantified by the subsequent ImageJ analysis (**m and o**; n=3 independent experiments). After measurement, F-actin was normalized by G-actin.

p, Coomassie Brilliant Blue (CBB) staining of the purified recombinant proteins. Each recombinant protein was expressed in *E. coli* and purified, followed by GST cleavage. Experiment was performed once.

q, Inhibition of interaction between CAPZs and actin by CRAD. Co-IP assay using purified recombinant proteins (see [Fig. 2m](#)) was quantified by ImageJ. n=3 independent experiments)

r and s. No effect of A511V-CRAD on Wnt/β-catenin signaling. AXIN2 qRT-PCR (**r**; n=independent experiments) and IB (**s**) using WT-CRAD or A511V-CRAD transfected CRC cells. IB was performed three times with similar results.

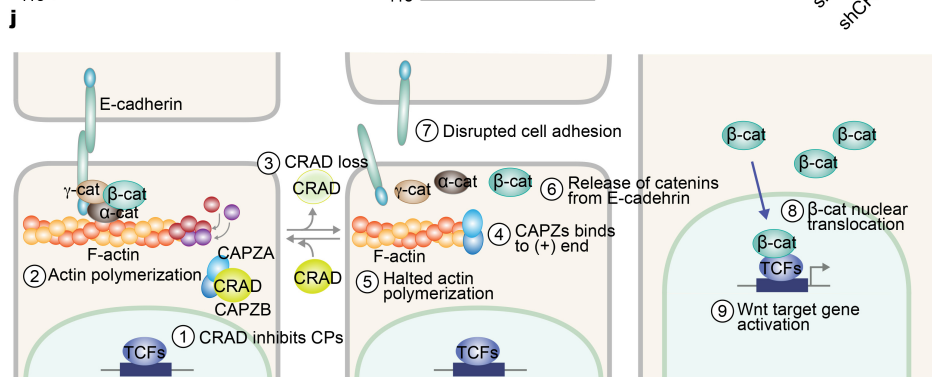
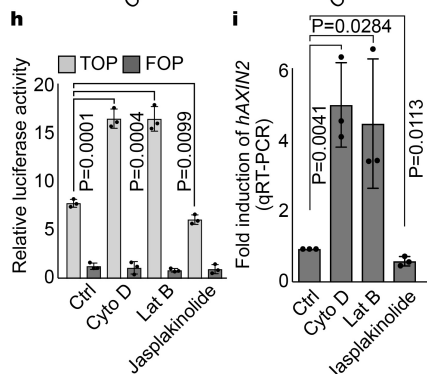
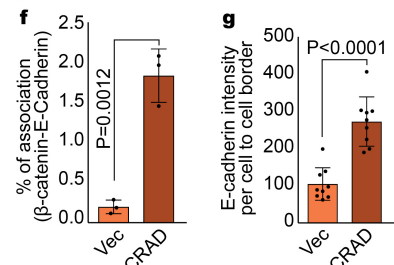
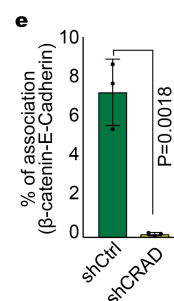
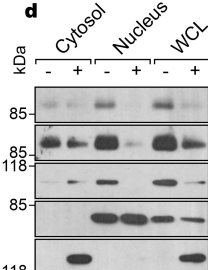
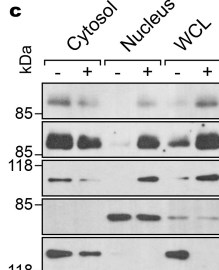
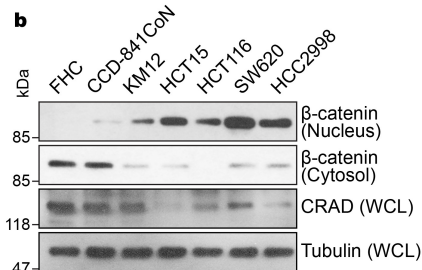
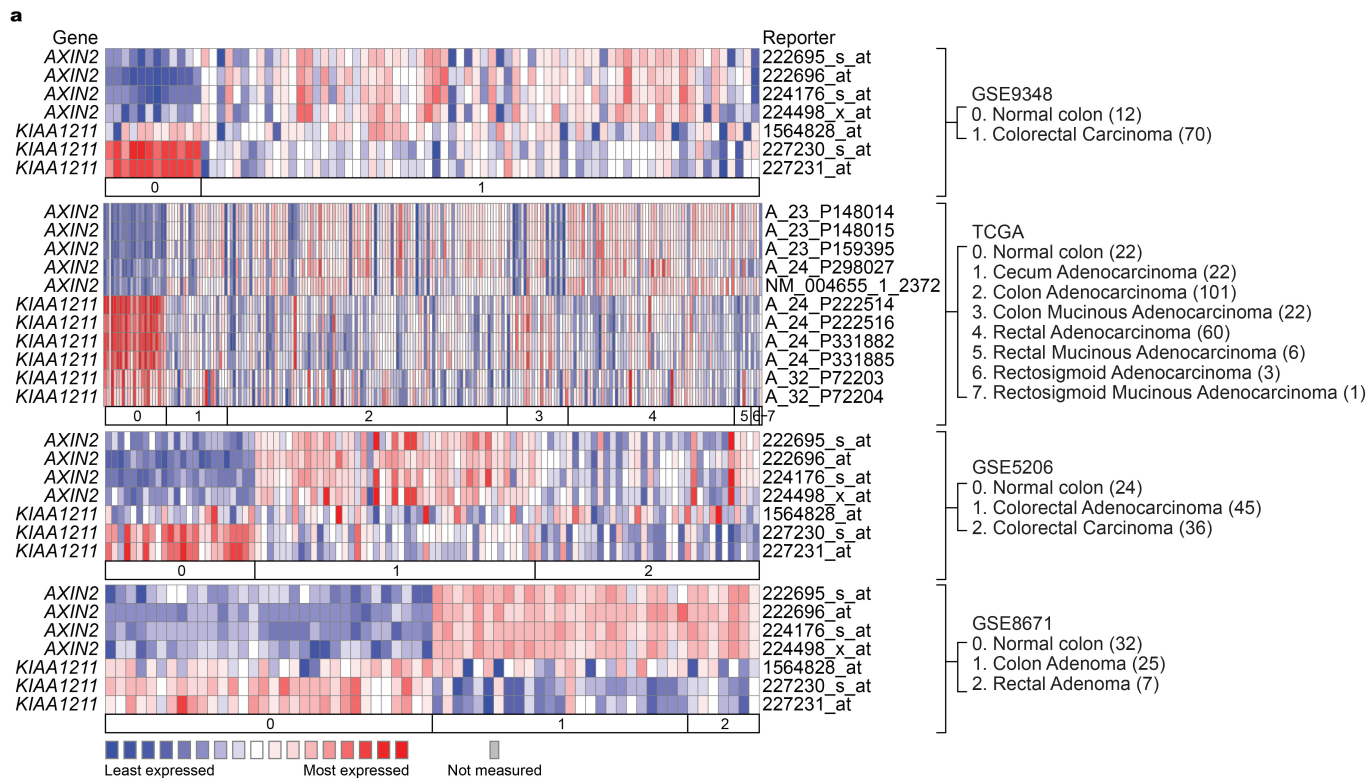
Representative images are shown; Scale bars indicate 20µm; Error bars: mean ± S.D.; NS: not significant (P>0.05); Two-sided unpaired *t*-test.



Supplementary Figure 3

Illustration of the working model of CRAD-induced F-actin polymerization.

CRAD inhibits CPs' binding to the barbed (+) end, resulting in the increased actin polymerization.



Supplementary Figure 4

CRC cell growth inhibition by CRAD

a, The mutually exclusive expression between *CRAD* and *AXIN2* in IECs and CRC. Oncomine analysis of *CRAD* and *AXIN2* expression in CRC.

b, Inversed correlation between CRAD and β -catenin. IECs and CRC cells were fractionated into the nucleus and cytosol fractions using fractionation kit. WCL: whole cell lysates.

c and d, Decreased nuclear β -catenin by CRAD. IECs (**c**) and CRC cells (**d**) were transfected with shCtrl or shCRAD and Vec or CRAD, respectively. After 48hr, cells were fractionated into the cytosolic and nucleus fractions, followed by IB.

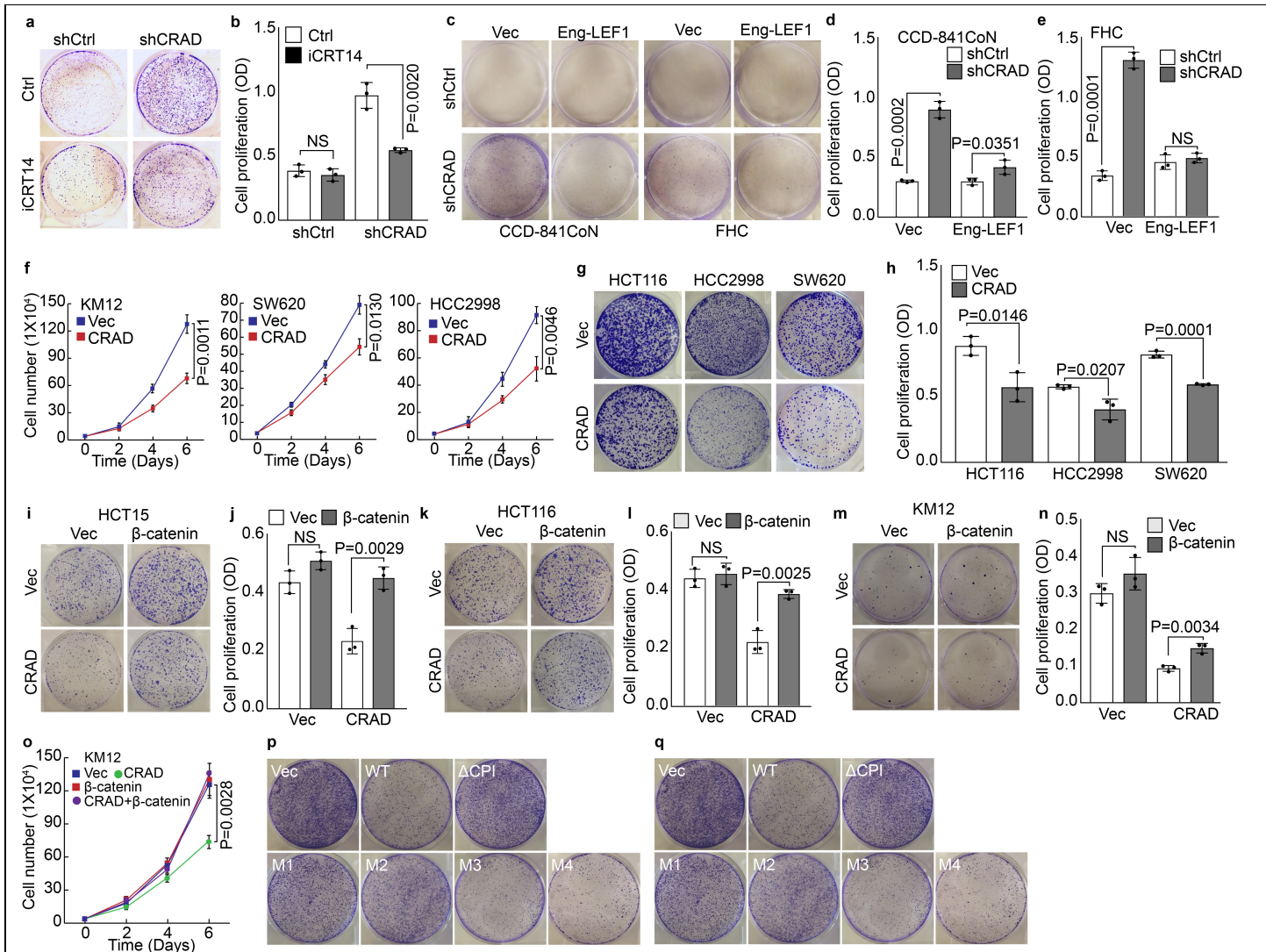
e and f, Increased interaction between E-Cadherin and β -catenin by CRAD. Cell extracts from IEC (**e**) and CRC cell (**f**) were used for IP analysis. Binding between E-Cadherin and β -catenin was quantified by ImageJ. n=3.

g, Increased interaction between E-cadherin and catenins by CRAD. HCT116 cells were transiently transfected with FLAG-CRAD plasmid and analyzed for IF staining using Super Resolution microscope (see Fig. 3p). E-cadherin intensity was quantified by ZEN software (Zeiss). n=10.

h and i, Blockade of the actin cytoskeleton increases Wnt/ β -catenin reporter activity. CCD-841CoN cells were transfected with β -catenin reporter plasmids (pMEGA-TOP/FOP-FLASH). 24hr after, cells were treated with Cytochalasin D (Cyto D; 2 μ M; barbed [+] end inhibition)⁵¹, Latrunculin B (Lat B; 1 μ M; monomeric G-actin inhibition)⁵², and Jasplakinolide (2 μ M; a stabilizer of actin cytoskeleton) for 72hr. β -catenin transcription activity was measured by TOP/FOP luciferase activity (**h**). CCD-841CoN cells were treated with indicated reagents as same concentration of (**h**). 72hr after treatment, cells were collected for qRT-PCR (**i**). n=3 independent experiments.

j, Illustration of the working model: the molecular mechanism of CRAD loss-induced hyperactivation of Wnt/ β -catenin signaling. In normal epithelial cells, CRAD inhibits CPs, which results in actin polymerization. Subsequently, increased F-actin stabilizes complex formation composed of E-cadherin-catenins-F-actin. In the absence of CRAD, F-actin is disrupted by CPs, which leads to destabilization of E-cadherin-catenin complex. Disturbance of E-cadherin-catenin complex releases β -catenin into the cytosols and the nucleus, which transactivates Wnt/ β -catenin target genes.

Error bars: average \pm S.D.; Two-sided unpaired *t*-test; Centre: Average.



Supplementary Figure 5

CRC cell growth inhibition by CRAD

a and b, Suppression of shCRAD-induced cell hyperproliferation by β-catenin inhibition in IECs. FHC (shCtrl and shCRAD) cells were treated with iCRT14 for 14 days and stained with crystal violet (**a**). Cell proliferation was quantified by measurement of absorbance at 590nm (**b**; n=3 independent experiments).

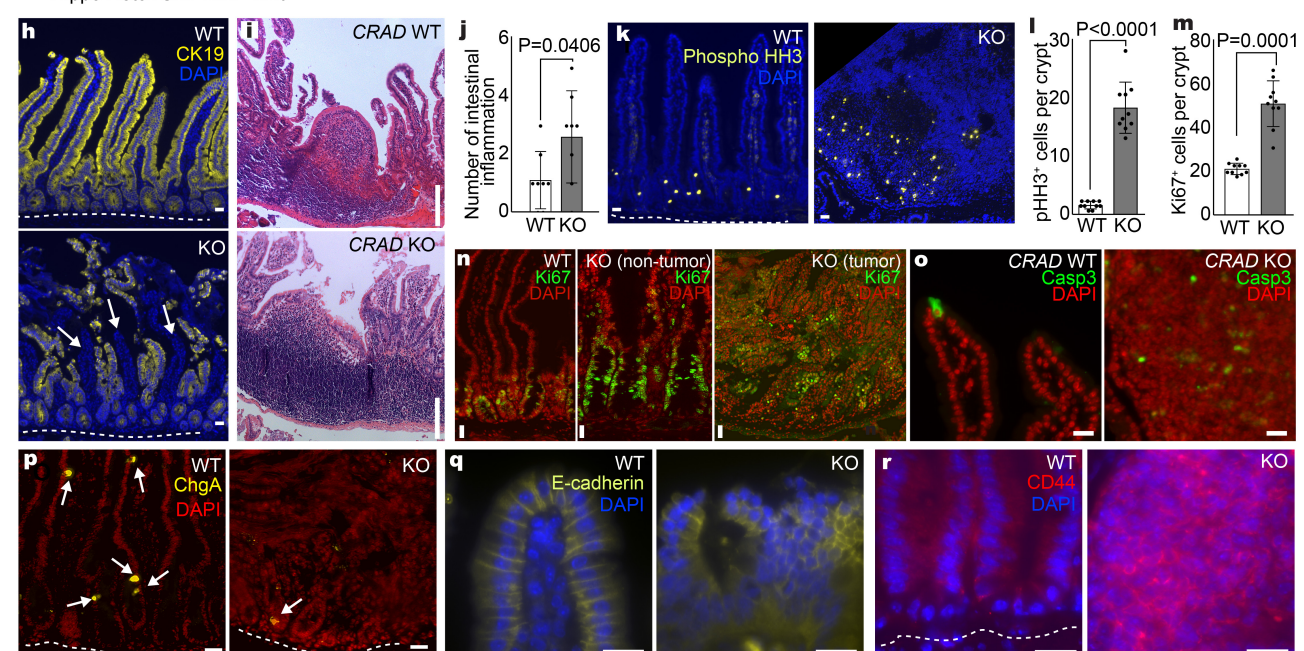
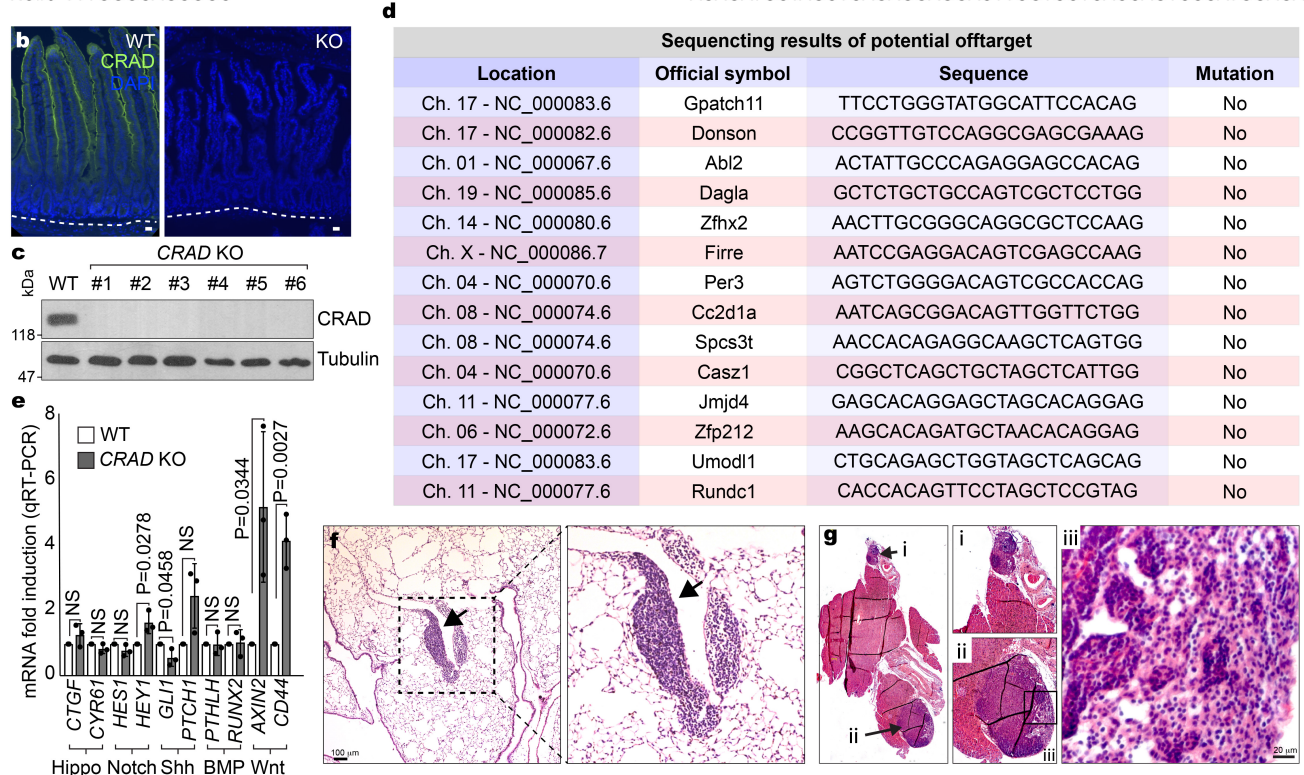
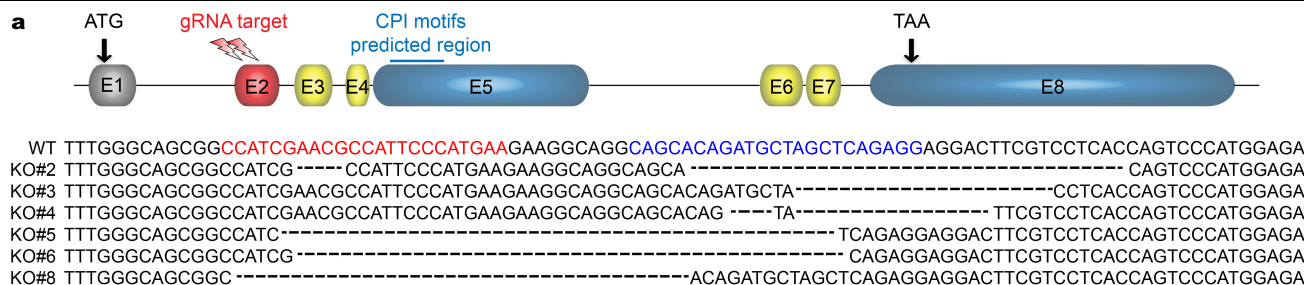
c-e, Engrailed-LEF1 inhibits *CRAD* depletion-induced IEC hyperproliferation. 1,000 cells of CCD-841CoN were seeded onto tissue culture plate. Cells were transfected with indicated plasmids. Eng-LEF1 was used as a dominant-negative mutant blocker of β-catenin-mediated gene transactivation^{53, 54}. 48hr after transfection, cells were incubated in 37°C tissue culture CO₂ chamber for 14 days. Colony forming was visualized by crystal violet staining (**c**) and quantified (**d and e**; n=3 independent experiments).

f-h, CRC cell growth inhibition by CRAD expression. CRC cells (Vec [control] and CRAD expressing) were analyzed for cell counting (**f**; n=3 independent experiments), crystal violet staining (**g**), and quantification (**h**).

i-o, β-catenin rescues CRAD-induced CRC cell growth inhibition. CRC cells were transfected with CRAD or-catenin plasmids and analyzed for crystal violet staining (**i,k,m**), quantification (**j, l, n**), and cell counting (**o**).

p and q, CRC cell growth inhibition by CPI motif-containing CRAD mutants. CRAD (FL, Δ CPI, and M1-M4)-transfected CRC cells were analyzed for quantification of cell proliferation. HCT116 (**p**); SW620 cells (**q**).

Representative images are shown; Error bars: mean \pm S.D.; NS: not significant ($P>0.05$); Two-sided unpaired *t*-test.



Supplementary Figure 6

Intestinal adenoma development by *CRAD* KO

a, CRISPR/Cas9-mediated targeting of *CRAD* alleles. Exon2 of *CRAD* was targeted using gRNAs. gRNA and Cas9 mRNA were injected into the pronuclei of C57BL/B6 mouse embryos.

b, *CRAD* expression in the small intestine. IHC of mouse intestine.

c, Validation of *CRAD* KO. The cell lysates were extracted from the small intestine of six independent *CRAD* KO mice for IB.

d, Sequencing analysis of potential off-target genes of *CRAD* gRNAs. No mutations in 14 off-target genes were detected.

e, Increased Wnt signaling target genes in *CRAD* KO mouse. *CRAD* KO-induced tumors were analyzed for qRT-PCR of Hippo, Notch, Shh, BMP, and Wnt signaling pathway target genes. n=3 independent mice.

f and g, Tumor development in the lung and pancreas of *CRAD* KO mice. Development of the early lesion of SCLC. The multiple sites displaying the early lesion of SCLC (black arrows; **f**) and the pancreatic tumors (**g**; i and ii) were observed. Scale bars indicate 100μm (**f**) or 20μm (**g**).

h, Disruption of epithelial cell integrity. Cytokeratin 19 (CK19). Arrows: Villi not expressing CK19.

i and j, Analysis of inflammation in *CRAD* KO mouse. Small intestine samples of WT and *CRAD* KO mouse were stained with H&E (**i**; n=8 mice). Intestinal inflammation (*foci* size > 200μm) was assessed by counting (**j**). Scale bars indicate 200μm.

k-n, Cell hyperproliferation in *CRAD* KO small intestine. Phospho-histone H3 (**k**) and Ki67 (**n**) staining and quantification (**l and m**; n=10 crypts). Scale bars indicate 20μm.

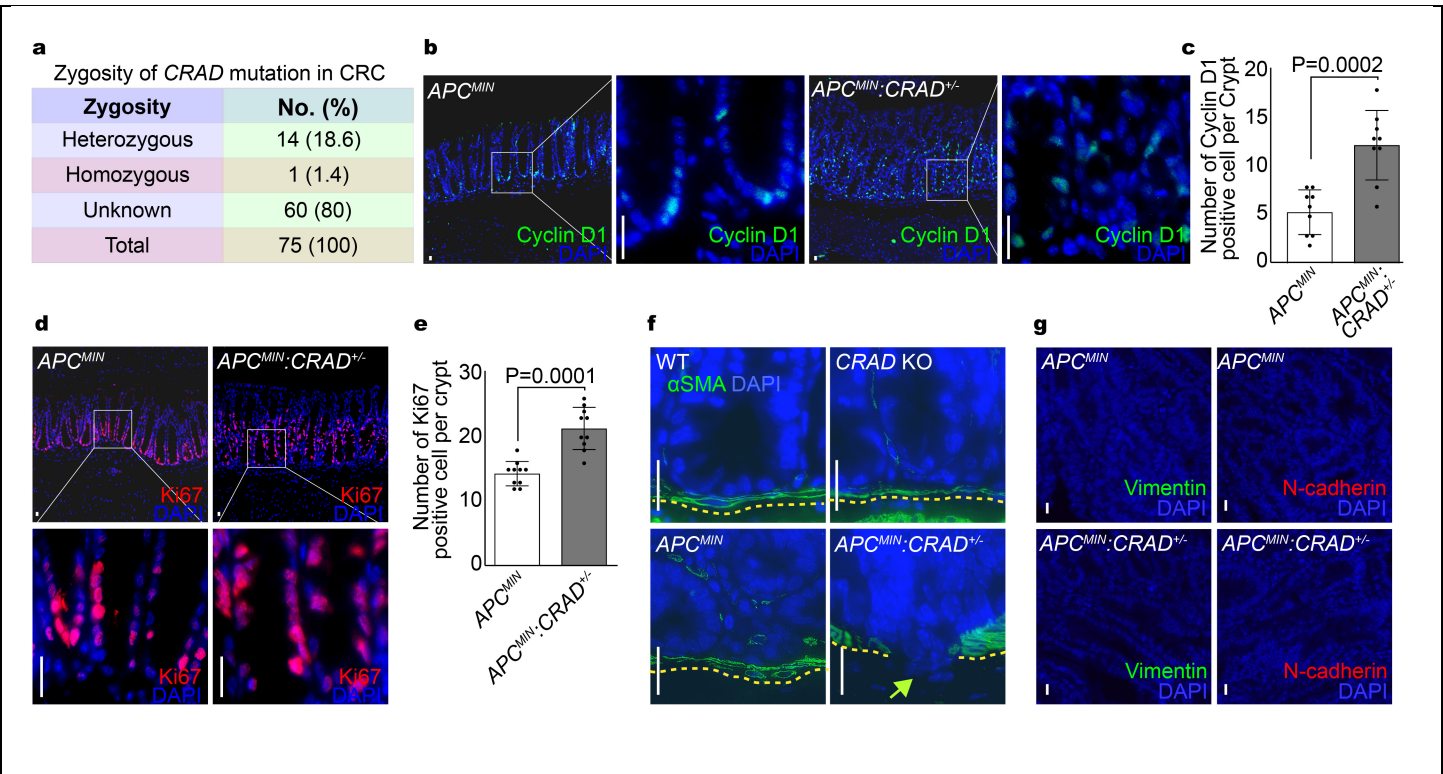
o, Analysis of apoptosis in *CRAD* KO-induced tumor lesion. WT and *CRAD* KO mice were analyzed for cleaved Caspase-3 (Casp3). Scale bars indicate 20μm.

p, Abnormal differentiation of IECs by *CRAD* KO. WT and *CRAD* KO small intestine were immunostained with ChgA (arrows). Scale bars indicate 20μm.

q, Disorganized cell adhesion in *CRAD* KO mice. Cells were stained with E-cadherin. Scale bars indicate 20μm.

r, Increased β-catenin target genes in the intestinal adenoma of *CRAD* KO mice. IHC for CD44. Scale bars indicate 20μm.

Representative images of three independent experiments; Error bars: mean ± S.D.; NS: not significant (P>0.05); Two-sided unpaired *t*-test.



Supplementary Figure 7

Accelerated intestinal tumorigenesis by *CRAD* heterogeneous KO

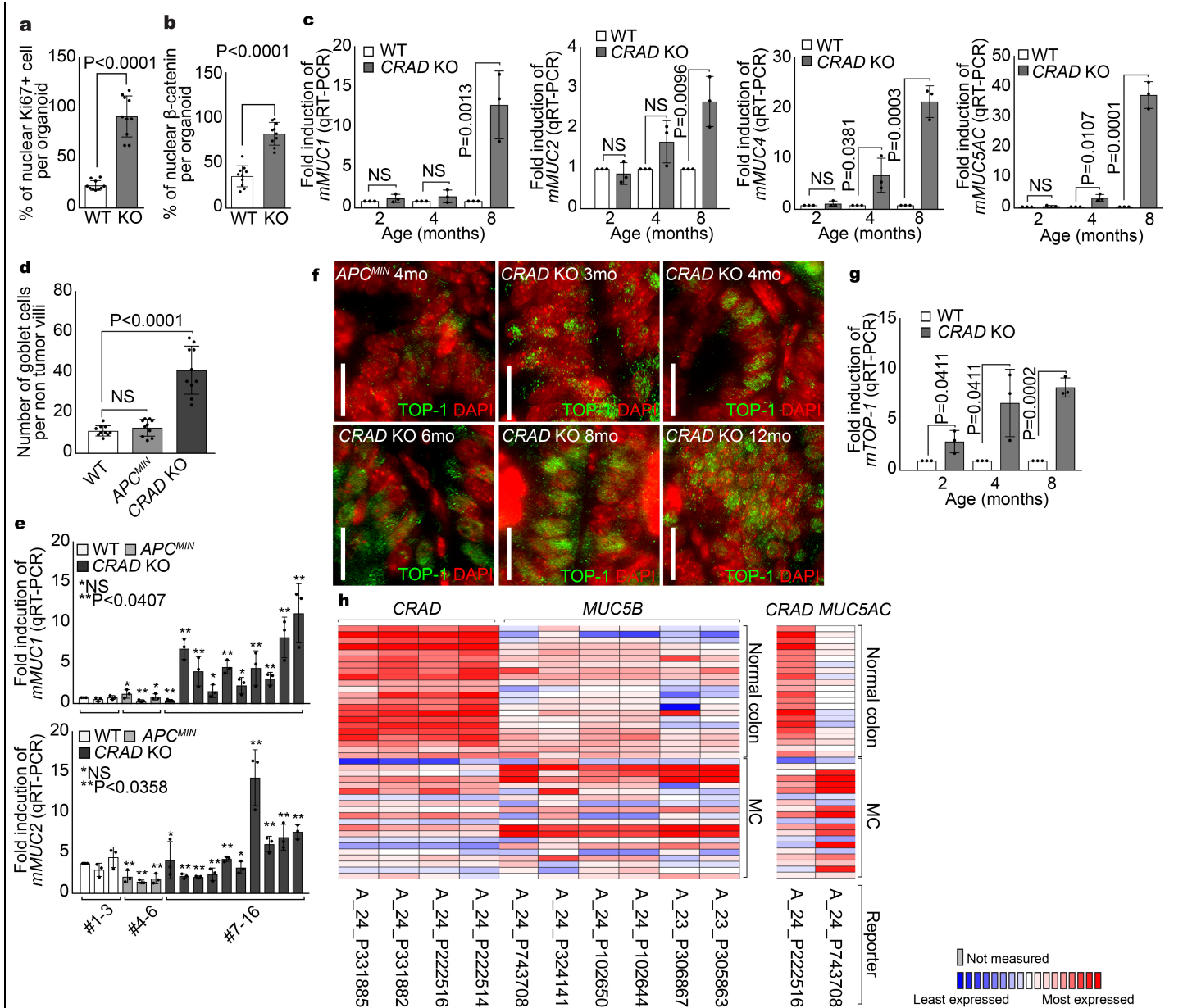
a, Heterozygous mutation of *CRAD* gene in CRC. Zygoty analysis of *CRAD* mutation in CRC patient samples using COSMIC and cBioportal databases. Of note, the frequency of heterozygous mutation is higher than that of homozygous mutation.

b-e, IHC of the non-tumor region of the colorectum of *APC^{MIN}* and *APC^{MIN}:CRAD^{+/-}* mice (4mo of age). Cyclin D1 (**b**); Ki67 (**d**). Quantification (**c**; n=9 crypts, **e**; n=10 crypts).

f, Micro-invasion by *CRAD* KO. Micro-invasion was observed in the tumor region of *APC^{MIN}:CRAD^{+/-}* small intestine. The small intestine samples were stained with α SMA (α -Smooth Muscle Actin: green) to visualize the basement membrane. The basement membrane of *APC^{MIN}:CRAD^{+/-}* is disrupted/discontinued tumor cell infiltration. Arrow indicates invasive tumor cell and yellow dashed lines mark the border of the basement membrane.

g, No EMT in tumor region of *APC^{MIN}:CRAD^{+/-}* mice. EMT marker analysis in *APC^{MIN}* and *APC^{MIN}:CRAD^{+/-}* small intestine samples. Markers of mesenchymal cell (Vimentin: green; N-cadherin: red) were not detected in tumors of both strains.

Representative images of three independent experiments; Scale bars indicate 20 μ m; Error bars: mean \pm S.D.; Two-sided unpaired *t*-test.



Supplementary Figure 8

Accelerated intestinal tumorigenesis by CRAD KO

a and b, IHC analysis of the organoids derived from CRAD WT and KO mouse intestine. Compared to WT, CRAD KO-derived cystic spheroids showed the increased cell proliferation (Ki67; **a**) and the increase of β -catenin (**b**). Quantification was performed using 10 cystic spheroids. $n = 10$ organoids from three different experiments.

c, Increased Mucin expression in CRAD KO. The small intestine samples from CRAD WT (2mo) and CRAD KO mice (from different ages as indicated) were examined by qRT-PCR for $mMUC1$, $mMUC2$, $mMUC4$, and $mMUC5AC$. $n = 3$ independent experiments.

d, Increased goblet cell in CRAD KO mice. After fixation and paraffin embedding, each sample was stained with PAS and quantified. $n = 10$ villi.

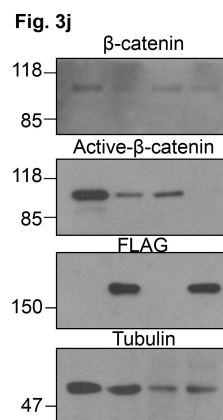
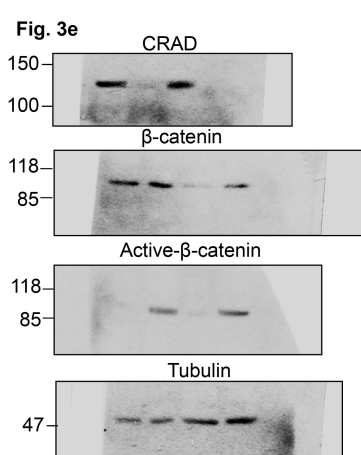
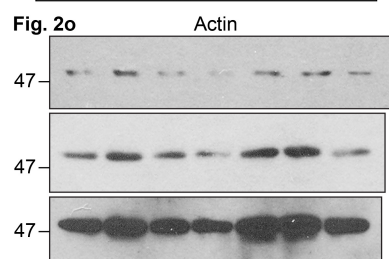
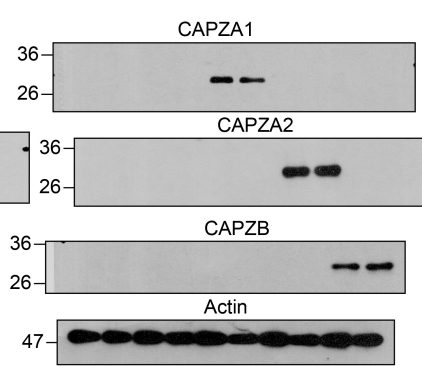
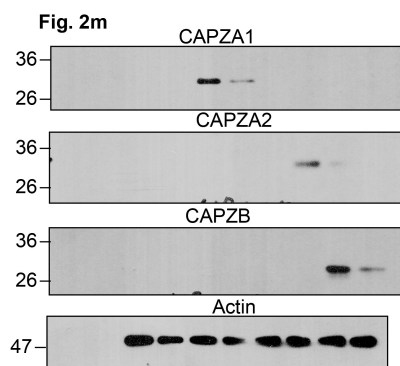
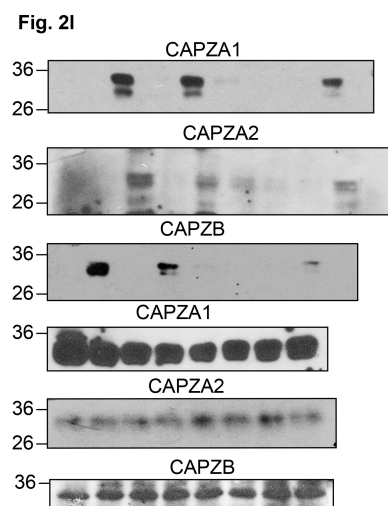
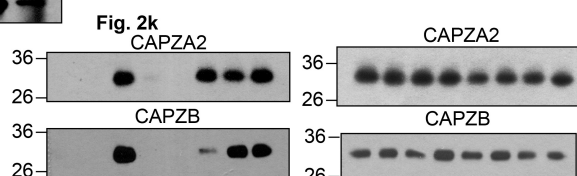
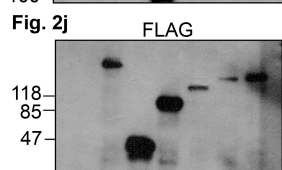
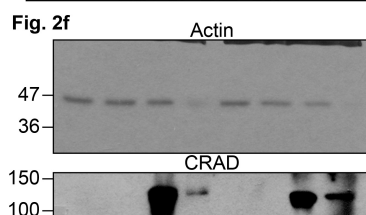
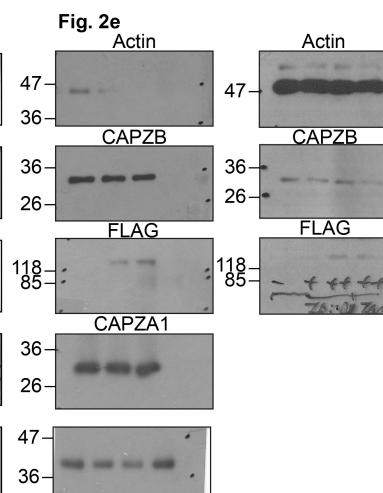
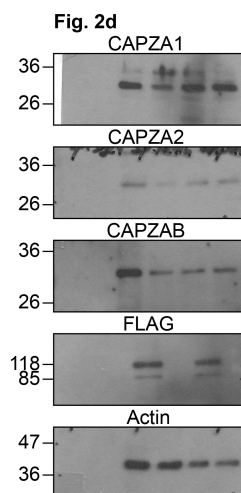
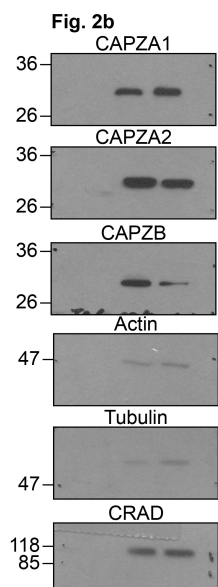
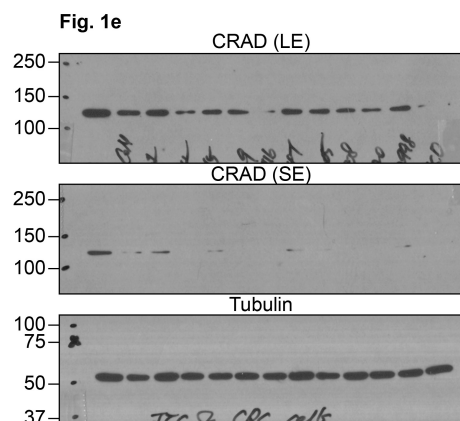
e, Increased mucin deposition in *CRAD* KO-induced tumors. qRT-PCR of *MUC1* and *MUC2* from the small intestine samples of each mice. n=3 independent experiments.

f, The increase of TOP-1 expression in *CRAD* KO mouse. Tumor of *APC^{MIN}* (4mo) and *CRAD* KO (3, 4, 6, 8, and 12mo) were immunostained with a TOP-1 antibody.

g, Upregulation of *TOP-1* in *CRAD* KO tumors. *CRAD* WT (2mo) intestine and *CRAD* KO tumors (from different age were examined by qRT-PCR of mTOP-1. n=3 independent experiments.

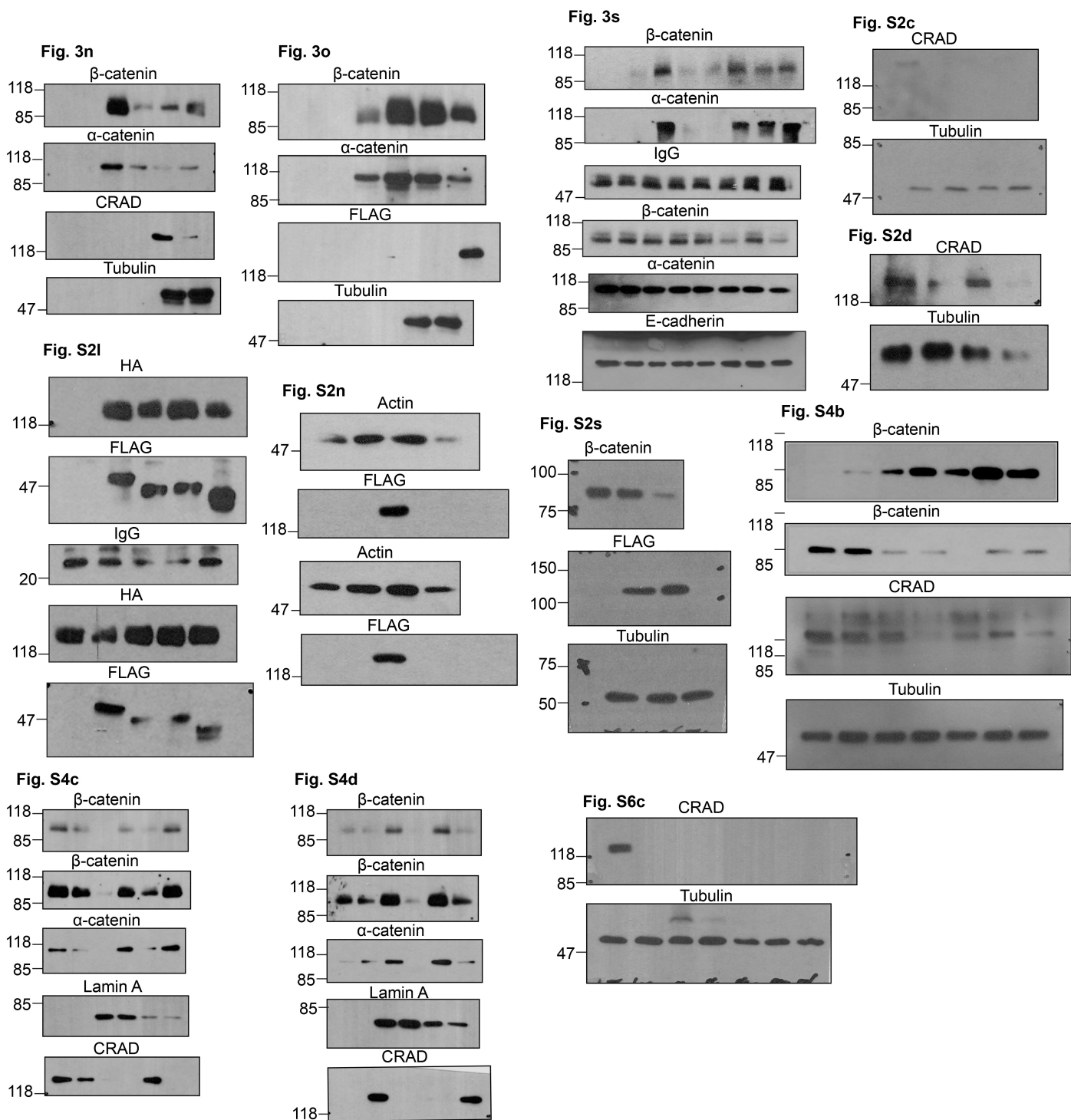
h, Mutual exclusive expression of *CRAD* and *MUCs*. Oncomine analysis of TCGA datasets; 10% gene rank; $P < 0.0001$; fold change > 2 ; compared with normal cells.

Representative images of three independent experiments; Scale bars indicate 20 μ m; Error bars: mean \pm S.D.; NS: not significant ($P > 0.05$); Two-sided unpaired *t*-test.



Supplementary Figure 9
Unprocessed blots
Figs. 1e-3j

SCANNED WB #2



Supplementary Figure 10

Unprocessed blots

Figs. 3n-S6c

Supplementary Table 1 Analysis of CRC tumor microarray for CRAD expression.

To determine CRAD protein expression, we analyzed 38 colon adenocarcinoma samples, 2 signet-ring cell carcinoma, and 14 normal colonic tissue samples using IHC for CRAD. While CRC samples show the low or absence of CRAD expression (colon adenocarcinoma: 12/44 [27%]; signet-ring cell carcinoma: 0/2 [0%]), normal tissue samples display the high expression of CRAD protein (14/14; 100%). NA: not applicable (due to the sample quality, these samples were excluded from analysis; 1: high expression; 0.5: low expression; 0: not detected).

Supplementary Table 2 CRAD-interacting proteins identified by tandem affinity purification and mass spectrometry.

293T cells stably expressing CRAD tagged with protein S, FLAG, Streptavidin-binding peptide (SFB) were processed for tandem affinity purification and mass spectrometry (TAP-MS/MS). Total 362 proteins were identified as CRAD-binding proteins. The number of peptides and peptide coverage are also included.

Supplementary Table 3 Information of MC microarray for CRAD expression.

To determine CRAD protein expression, we analyzed 34 MC and normal colonic tissue samples (Biomax; OD-CT-DgCol03-003 : Colon mucinous adenocarcinoma tissue array) using IHC for CRAD. While MC samples show the low or absence of CRAD expression normal tissue samples display the high expression of CRAD protein. TNM grading: T - Primary tumor; N - Regional lymph nodes; M - Distant metastasis.

Supplementary Table 4 Primer information.

A complete list of primers.

Supplementary Table 5 Antibody information.

A complete list of antibodies.

Supplementary Table 6 Statistics Source Data.

14 sheets contain statistics source data.

References

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